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Introduction

The most critical aspects of reliable food contamination analysis are the reduction of interferences from the sample matrix and analyte recovery. Traditionally, SPE, SLE, Liquid-Liquid, syringe filtration, and centrifugation have been used to reduce matrix interference prior to MS analysis. However, these techniques are time consuming, adversely impact recovery, require expensive consumables, and use large amounts of solvent (which then need to be concentrated). Several studies were undertaken to investigate different designs of filter vials to improve sample clean-up methods in orange juice, apple juice, grape juice, vegetable juice and water.

Thomson **extreme** Filter Vials (patented) offer multi-layer filtration for viscous samples and samples containing up to 30% solid particulates. The filter vial consists of two parts: a filter vial shell and a plunger, which includes the multi-layer filter on one end and a vial cap on the other end. Samples are filtered by pipetting the sample into the filter vial shell, inserting the plunger into the shell, and then pushing the plunger into the shell. The filtration process from sample pipetting to autosampler ready only requires 15 seconds. Benefits to the use of Thomson **extreme** Filter Vials include lower cost, faster sample preparation time, less use and disposal of organic solvents, and in some instances improved recoveries.

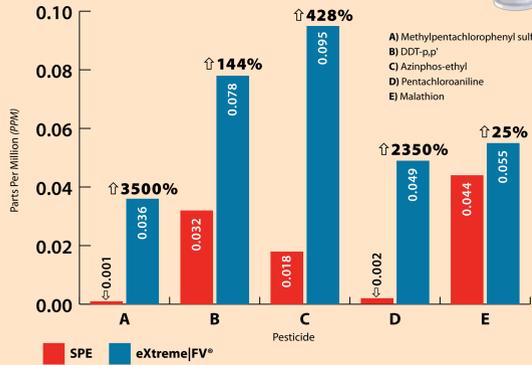
Thomson **exTractor3D** Filter Vial (patented) offer filtration with increased volume, enabling multiple extraction techniques with different resins/sorbents or solids/large particulates (greater than 35%) to autosampler ready vials. **exTractor3D** is a product uniquely designed for the addition of resins/sorbents, QUCHERS dispersive salts, pills, or special resins in the standard autosampler ready vial. The filter vial consists of two parts: a filter vial shell and a plunger which includes a multi-layer filter on one end and a screw cap on the other end. Large solids/large particulates can be placed within the **exTractor3D** where multiple extraction techniques occur. Prior to the introduction of the **exTractor3D**, samples required multiple steps using SPE or other methods to remove interfering analytes and co-eluting compounds. SPE or QUCHERS can now be completed with multi-step filtration without risk of solids compromising the autosampler. Pills and other large solids can be broken down for complete testing using the **exTractor3D**. The **exTractor3D** allows for compounds to be separated from the matrix with the addition of resins/sorbents, resulting in both a higher signal to noise ratio and peaks that are more differentiated.



Method I- extreme Filter Vials® vs SPE for the analysis of Pesticides in Orange Juice by GC/MS

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Comparison of Pesticide Recoveries



Abstract

Pesticides act as toxins when found in sufficient quantities as residues in food. This is of particular importance for orange juice because it is consumed in high quantities. Sensitive, rapid, and cost effective analytical methods are required in order to reduce the risk to consumers. Solid Phase Extraction (SPE) is a common sample preparation technique used prior to GC or LC analysis of pesticides in food. Typically, SPE is used to concentrate analytes, reduce interference from co-eluting molecules or to clean up "filter" sample particulates. Drawbacks to the use of SPE include cost, sample preparation time, large sample volumes, use and disposal of organic solvents, and potentially poor recoveries. The continuing development of higher sensitivity instrumentation and improved filtration devices has led many labs to investigate whether methods can be adapted to eliminate the SPE step.

Thomson **extreme** Filter Vials offer multi-layer filtration for viscous samples and samples containing up to 30% solid particulates. Filtration time from unfiltered sample transfer to filtered sample in an autosampler ready vial is only 15 seconds. The filter vial consists of two parts: a filter vial shell and a plunger which includes the multi-layer filter on one end and a vial cap on the other end. Samples are filtered by pipetting the sample into the filter vial shell, inserting the plunger into the shell, and then pushing the plunger into the shell.

Prior to the introduction of the **extreme** Filter Vials, many samples containing high levels of particulates were only "filtered" by using an SPE step in the method. These methods are readily amendable to the replacement of the SPE step with a much faster and lower cost **extreme** Filter Vial step.

Experiment

Samples were prepared and analyzed at Micro Quality Labs, Burbank, CA.

- Sample Extraction:**
- 1) Spike 10mL of commercially available High Pulp Orange Juice with 1mL of 1 ppm pesticide standard mix in a 40mL vial.
 - 2) Add one pack (approximately 6g) of Restek Extraction Salts (Restek catalog #26230) to the spiked orange juice.
 - 3) Extract the spiked orange juice with 4 x 25mL portions of methylene chloride.
 - 4) Concentrate to dryness using a Turbovap II concentrator.
 - 5) Dissolve the residue in approximately 10mL of acetonitrile.
 - 6) Vortex and sonicate the re-suspended residue with frequent swirling.
 - 7) Split the re-suspended residue into two 5mL portions.
 - 8) Dilute each 5mL portion with acetonitrile to 10mL using a volumetric flask.
 - 9) Label one flask "for SPE" and the other "for Thomson **extreme** Filter Vial".

SPE Cleanup Prior to Analysis - Restek 6mL Combo SPE Cartridge

- 1) Wash one Restek 6mL Combo SPE Cartridge (packed with 200mg CarboTrap 200 and 400mg FSA Restek catalog #26127) with acetonitrile.
- 2) Add the 10mL portion of the re-suspended residue from the flask labeled "for SPE" to the SPE cartridge.
- 3) Elute the sample from the cartridge with 50mL of acetonitrile.
- 4) Concentrate the eluted sample to 10mL using a Turbovap II concentrator.

Thomson extreme Filter Vial Cleanup Prior to Analysis

- 1) Add 40µL of the re-suspended residue from the flask labeled "for Thomson **extreme** Filter Vial" to the shell of one Thomson **extreme** Filter Vial 0.45µm PTFE (Thomson Part Number 85540-500).
- 2) Insert plunger completely.

Compound	Sample name	SPE - Residue	Syringe Filtration	ppm	extreme Filter Vial	ppm
Azinphos-methyl		0.028		0.035		
Azinphos-ethyl		0.023		0.115		
Bromophos-ethyl		0.025		0.057		
Cyfluthrin		0.082		0.113		
Cyhalothrin (trans-isomer)		0.026		0.099		
Cypermethrin I (Z/E)		0.082		0.117		
Cypermethrin II (CAS # 52135-07-8)		0.08		0.113		
Cypermethrin III (Beta)		0.058		0.104		
Cypermethrin IV (CAS # 52135-07-8)		0.07		0.097		
DDT-p,p'		0.035		0.065		
DDT-p,p'		0.032		0.078		
Deltamethrin		0.053		0.102		
Endosulfan (alpha isomer)		0.041		0.076		
Fenitrothion sulfone		0.081		0.107		
Fenvalerate I		0.076		0.106		
Fenvalerate II (CAS # 51830-58-1)		0.055		0.073		
Fluvalinate I (CAS # 102851-06-9)		0.058		0.084		
Methylparathion/phenyl sulfide		0.001		0.036		
Octachlorodipropyl ether (OADE)		0.021		0.047		
Permethrin		0.062		0.046		
Permethrin I		0.068		0.097		
Permethrin II (trans)		0.071		0.115		
Phosalone		0.005		0.089		
Phosmet		0.031		0.094		
Phothos		0.033		0.06		

Table 3: Pesticides in Orange Juice Comparison of SPE to extreme Filter Vials

Analysis

Samples were analyzed utilizing an Agilent Technologies GC/MS, 7000 Triple Quad system equipped with a 7890A GC system and 7693 auto sampler.

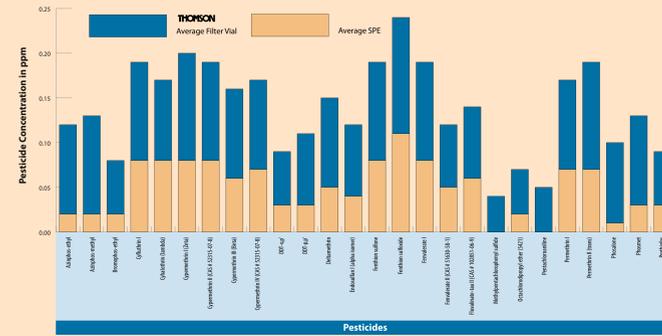
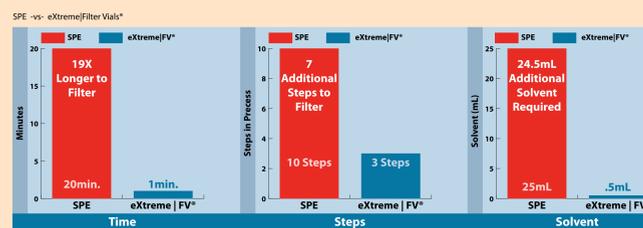


Fig 3: Pesticides in Orange Juice Comparison of SPE to extreme Filter Vials

What did we learn?



Conclusions

The Thomson **extreme** 0.45µm PTFE Filter Vials patented (Thomson #85540-500) yielded 26% higher recoveries on average when tested with 87 common pesticides. In the cases highlighted in the results table, greater than 428% recovery increases were seen. In the case of Hexachlorobenzene, no pesticide was detected in the sample prepared by SPE and 0.019 ppm was detected in the sample prepared with the **extreme** Filter Vial. The use of Thomson **extreme** 0.45µm PTFE Filter Vials as a substitute for SPE conforms to USP Method 561.

The results show Thomson **extreme** Filter Vials offer a viable alternative with higher recovery and less preparation time compared to SPE for the preparation of juices prior to pesticide analysis.

Method III- Screening and Quantitation of 250 Pesticides in Fruit Juices with Positive/Negative Switching LC/MS/MS

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Introduction:

LC/MS/MS operated in Multiple Reaction Monitoring (MRM) mode with dual scan Electrospray Ionization (ESI) is widely used for polar, semi-volatile, and thermally labile pesticides in food testing. Many labs currently perform multi-residue analysis of pesticides using separate positive and negative methods due to instrument limitations. This requires twice the sample and twice the analysis time. The Bruker EVOQ Elite LC-Triple Quadrupole System provides fast positive/negative switching allowing for simultaneous determination of positive and negative co-eluting compounds numbering in the hundreds. A study using the EVOQ for the analysis of 250 pesticides in apple juice, cranberry juice, grape juice, orange juice and vegetable juice using only one method with positive/negative switching for over 500 MRM transitions. The measurements were conducted using store bought juices with a dilute-and-shoot approach without sample enrichment.

Sample Preparation:

1. Mix 50µL fruit juice with 450µL of solvent (MeOH:water, 10:90, v/v) directly into the outer shell Thomson **extreme** Filter Vial, 0.2µm PVDF and press filter plunger to filter
- * Store bought Fruit include: Apple Juice, Cranberry Juice, Grape Juice, Orange Juice and Vegetable Juice
2. Load the Thomson **extreme** Filter Vials into the autosampler

Equipment:

- EVOQ Elite Triple Quadrupole Mass Spectrometer
- Bruker UHPLC
- CTC Autosampler
- Source: HESI
- Spray Voltage Positive: 4000V
- Spray Voltage Negative: 4000V
- Column: YMC-Pack ODS-AQ 3µm
- Column Temperature: 40 °C
- Injection Volume: 30 µL
- Mobile Phase Gradient: 5 mM Ammonium Fluoride in Water/Methanol
- Flow rate: 400 µL/min

Results:

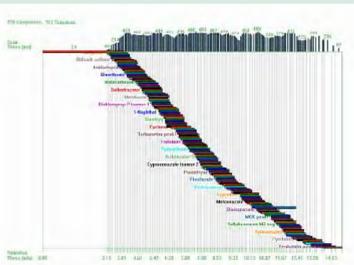


Fig 1. The compound based scanning (CBS) of 250 pesticides can automatically compute and assign the scan (dwell) time for each MRM for timed MRM, based on peak width and data points required.

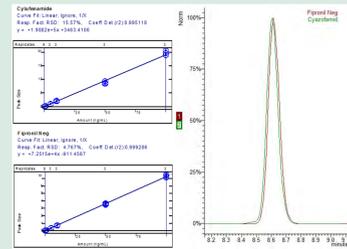


Fig 3. Calibration curve of negative pesticide Fipronil (left top) and positive pesticide Cyazofamid (left bottom), and their co-eluting plots (right).

Fruit Juice=>	Apple Juice	Orange Juice	Cranberry Juice	White Grape Juice	Vegetable Juice
Pesticide			µg/L (ppb)		
Azinrotrilin	ND	ND	0.32	ND	0.48
Boscalid	ND	0.16	ND	ND	ND
Carbaryl	ND	0.39	1.47	ND	ND
Carbofuran	ND	0.14	ND	ND	ND
Dimethoate	ND	0.30	ND	ND	ND
Imidacloprid	ND	ND	0.60	ND	0.20
Mandipropamid	ND	ND	0.59	ND	ND
Metasuliy	ND	ND	0.21	ND	ND
Methoxyfenozid	ND	ND	ND	ND	0.84
Tebuconazole	ND	ND	0.32	ND	ND
Thiabendazole	1.8	ND	ND	ND	ND

Table 1. Test results for store bought juices. ND is not detected <0.1ppb

Conclusion:

- The calibration on triplicate injections showed excellent linearity and response factor RSD over 3 orders range using a simple sample prep of adding juice and water directly into the Thomson **extreme** Filter Vial and then to the autosampler.
- Good linearity, sensitivity and response factor RSD for positive and negative co-eluting pesticides.
- A total of twelve pesticides were detected in apple juice, orange juice, cranberry juice or vegetable juice
- The multiple pesticides detected in orange juice, cranberry juice and juice may suggest that juice comes from multiple sources of raw materials or pooled juices.



TIC Chromatograms

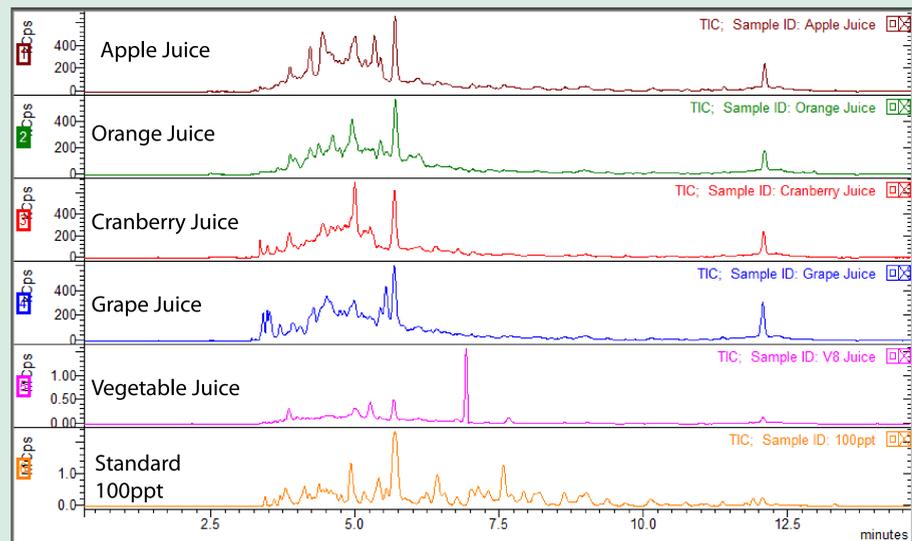
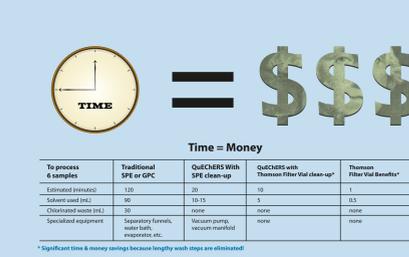


Fig 4. Total Ion Chromatograms (from top to bottom): Apple juice, orange juice, cranberry juice, grape juice and vegetable juice.

Overall Conclusion



Final Conclusion

The methods presented here for the analysis of contaminants in juices and water show the Thomson Filter Vials compared to traditional methods of clean-up, including SPE, liquid-liquid extraction and syringe filtration.

For sample clean-up, post extraction, in the analysis of pesticides of orange juice by GC/MS, the Thomson **extreme** Filter Vials, showed improved recovery of many of the pesticides by as much as 428%. The improved method yielded higher recovery and, used less solvents and less sample preparation time by eliminating the SPE step for clean-up for the analysis of pesticides of orange juice. Simply add the extracted sample to the Thomson **extreme** Filter Vials, PTFE 0.45µm (P/N 85540-500), depress the plunger to filter and load onto the autosampler.

Analyzing 250 pesticides in store bought apple, grape, orange and vegetable juices by LC/MS sample preparation was streamlined to diluting the juice directly into the **extreme** Filter Vials, 0.2µm (P/N 85531-500) and placing the filter vial onto the HPLC. This method has excellent linearity and sensitivity down to <0.1ppb in real life samples.

In the analysis of Hexavalent Chromium, the Thomson Standard Filter Vials, PTFE 0.45 µm (P/N 35540-500) showed no chromium contamination in the vial or membrane materials and reproducible analysis at 0.1ppb. Thomson Standard Filter Vials replaced 4 part numbers: syringe, syringe filter, autosampler vial and cap and saves hours of technician time pre-cleaning vials.

What do all these methods have in common? All of the Thomson Filter Vials simplify sample preparation, lower costs and save time.

Thomson Instrument Company is not affiliated with Bruker®, Micro Quality Labs, Canadian Ministry of Environment, YMC, Agilent, Restek or their products.

Method II- The Determination of Hexavalent Chromium in Waters by Ion Exchange Chromatography-Inductively Coupled Plasma Mass Spectrometry (IC-ICP-MS)

This method utilizes a hyphenated technique, i.e. ion exchange chromatography (IC) coupled to an inductively couple plasma mass spectrometry (ICP-MS) to determine Cr(VI) in treated drinking water, surface water and ground water. Samples are collected and preserved at a pH > 9 condition, and then injected directly into an anion exchange column. Cr(VI) is separated from other possible Cr species and other metals by the anion exchange functioning group inside the column. The column eluent is introduced directly into the sample introduction interface and the ionization source of the ICP-MS. Chromium chromatographic peak is identified and quantified by the mass spectrometry with external calibration.

Previous Labware Cleaning Procedure:

It is critical to pre-clean and dry labware in a clean flow bench, in order to minimize contamination.

- Place tubes and caps into 10% Nitric Acid (made from reagent grade) acid bath for at least 24 hours
- Transfer tubes and caps into a DI Water bath to soak for at least 24 hours
- Remove tubes and caps, rinse with DI Water at least three times
- Remove as much water as possible and place inside a Class 10 Vertical Laminar Flow Metal Free Hood and let dry

Sample Requirements:

- Sample must be preserved to achieve pH > 9 with Ultra Pure Concentrated Ammonium Hydroxide
- Sample is collected in a 15mL amber high density polyethylene (HDPE) bottle with a plastic cap
- Samples are stored at < 8°C for up to 30 days, provided that the sample containers are sealed properly and stored in an acid fume free environment. However, it is recommended that samples be analyzed as soon as possible upon receipt.

Sample Preparation:

1. Check sample pH using a pH testing strip by transferring a small volume of sample to prevent cross contamination. If the pH is > 9, sample is ready for IC-ICP-MS analysis.
2. Label the Thomson 0.45 µm PTFE Filter Vials (35540-500).
3. Pipette 0.5mL of the sample into the filter vial shell.
4. Partially insert the filter vial plunger into the filter vial shell.
5. Place filter vials in the Thomson Toggle Press and press the lever to filter the samples (can press up to 5 vials each time).
6. Load the filter vials into the Varian autosampler.
7. Include Calibration Standards (0.05 µg/L, 0.1 µg/L, 0.5 µg/L, 1.0 µg/L) and QC Standards (DI Water Blank, Tap Water Blank, Tap Water Spiked) for every 20 samples analyzed.

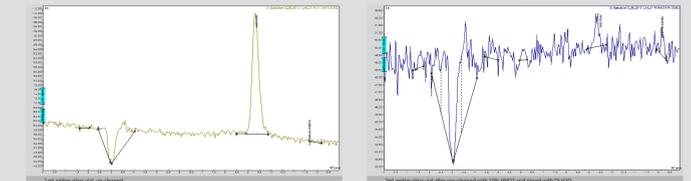
Note: r² > 0.995 for the calibration curve

Equipment:

- Varian ProStar 210 HPLC
 - Varian 820MS ICP-MS
 - Pump Rate (rpm): 20
 - Stabilization delays: 0
 - Skimmer Gas Source: H₂
 - Skimmer Flow: 30
 - Column: Hamilton PRP-X100 Anion Exchange Column & Guard Column
 - Mobile Phase:
 - Mobile Phase A: 100mM Ammonium Nitrate, pH > 9, pH adjust with 16N Nitric Acid
 - Mobile Phase B: DI Water, pH > 9, pH adjust with Ultra Pure Ammonium Hydroxide
- | Time | Flow (mL/min) | %A | %B |
|---------|---------------|----|----|
| Pre-run | 1.0 | 80 | 20 |
| 9.0 | 1.0 | 80 | 20 |



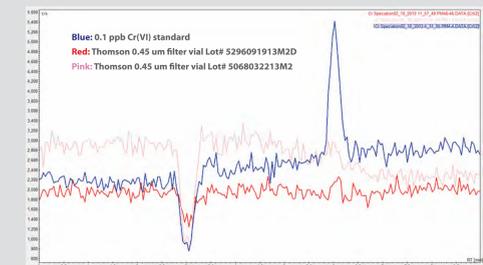
Existing Canadian HEXCR-E3510 Procedure used a Syringe & Filter, amber glass vial and cap to filter water samples prior to analysis. This required pre-cleaning 110% HNO₃, rinse with DI-H₂O and dried of the amber glass vials



Results:

Results of spiked hexavalent chromium calibration standard in the concentrations of 0.05µg/L, 0.1µg/L, 0.5µg/L, 1.0µg/L yielded r² > 0.995. Figure 1 shows a chromatogram of a 0.1ppb Cr(VI) standard overlaid with 2 different lots of blank samples.

Figure 1: Chromatogram of a 0.1ppb Cr(VI) standard overlaid with 2 different lots of blank samples



Conclusions:

The Thomson Standard Filter Vials showed no chromium contamination in either the filter vial materials or the filter membrane allowing for their use in the analysis of hexavalent chromium in water. The validation of the Thomson Standard Filter Vials into a validated method for the analysis of hexavalent chromium reduces both time and waste compared to the previous filtration method.



Filter Vial
PTFE 0.45µm
35540