

Improved Method for the Analysis of a Pain Management Supplemental Panel in Urine using the Thomson eXtreme Filter Vials® by LC-MS/MS

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Introduction or Abstract

This improved sample preparation method allows for the quantitative measurement of the following pain management drugs in urine. The urine samples were diluted and filtered using Thomson eXtreme|FV®, followed by LC/MS/MS analysis. The most critical aspects of reliable urine analysis are the reduction of interferences from the sample matrix and analyte recovery. Traditionally, SPE, SLE and centrifugation have been used to reduce matrix interference prior to MS analysis. However, these techniques are time consuming, adversely impact recovery, require expensive consumables, lab equipment and use large amounts of solvent. Thomson eXtreme|FV® (patented) offer multi-layer filtration for viscous samples and samples containing up to 30% solid particulates. The filter vial consists of two parts: a filter vial outer shell and a plunger, which includes the multi-layer filter on one end and a vial cap on the other end.

Experimental

Equipment:

ABI 4500 Mass Spectrometer

Shimadzu Prominence HPLC equipped with

Autosampler: SIL-20AC HT
 Pumps A, B: LC-20AD
 Communication Bus Module: CBM-20A
 Column Oven: CTO-20A
 Degasser: DGU-20A_R
 Column: Ultra Biphenyl Columns (5µm 50 x 2.1 mm) - Restek

Method:

Flow Rate: 0.5 mL/min

Mobile Phases:

A: 0.1% Formic Acid in HPLC Water
 B: 0.1% Formic Acid in Methanol
 Run Time: 8.5 minutes
 Injection Volume: 15µL

Eppendorf Mix Mate Vortex Mixer

Thomson eXtreme|FV® 0.2µm PVDF (p/n 85531)

Thomson 48 position Vial Filter Press (p/n 35010)

Improved Sample Preparation Post Hydrolysis

- Place 400 µL of 20% MeOH / 80% Water / 0.1% Formic Acid in each of the outer shells of the Thomson Filter Vials
- Add 25µL of Standard/Control/Patient Sample + 10µL of Internal Standard
- Place Thomson Filter Plunger on top of the Thomson vial, Thomson vials -eXtreme|FV® 0.2µm PVDF, w/ Pre-Slit Red Cap #85531.
- Press filter plunger down approximately ¼ of the way into each of the Thomson vials.
- Vortex for 30-40 seconds
- Slowly press filter plunger the rest of the way down using the Vial Filter Press.
- Extracts are ready for LC/MS/MS analysis using the Shimadzu / ABI 4500
- Inject 15µL

Results

This improved sample preparation method allows for the quantitative measurement of the following pain management drugs in urine, Table 1. The improved method utilizes the Thomson eXtreme|FV® for sample clean-up significantly reducing the cost and time of per sample analysis. This method was validated for all 17 drugs in the supplemental pain management panel over 3 days.

Table 1. Drugs analyzed as part of the Pain Management Supplemental Panel in urine

| Drug | Drug | Drug | Drug | Drug |
|----------------|----------------|-----------------|----------------------|----------|
| Amirtriptiline | Cycloenzaprine | Desipramine | Ritalinic Acid | Tramadol |
| Nortriptyline | Duloxetine | Meperidine | Pregabalin | |
| Carisoprodol | Gabapentin | Normeperidine | Tapentadol | |
| Meprobamate | Imipramine | Methylphenidate | Tapentadol-O-Sulfate | |

Data

Fig.1. Mass spectrum of the 17 drugs included in the Supplemental Pain Management Panel in Urine.

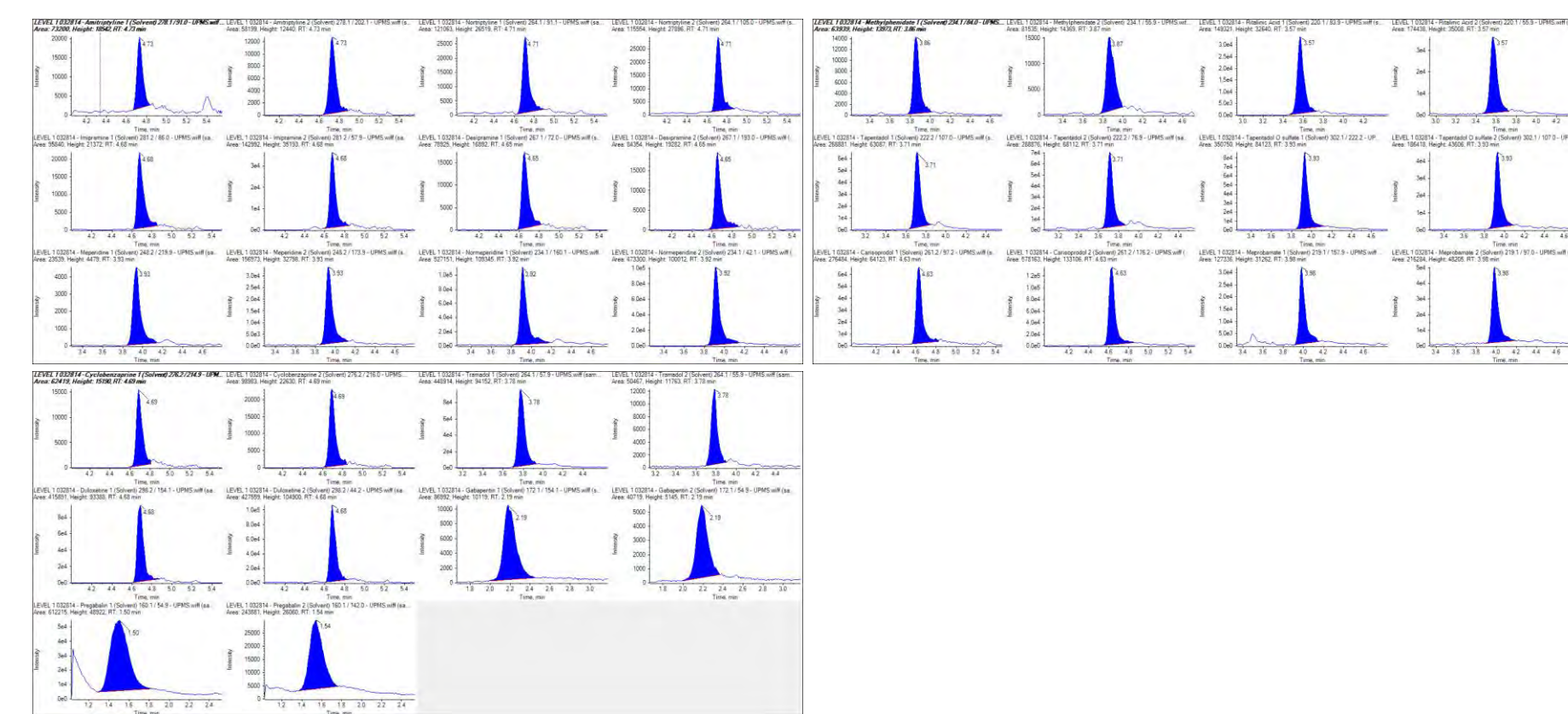
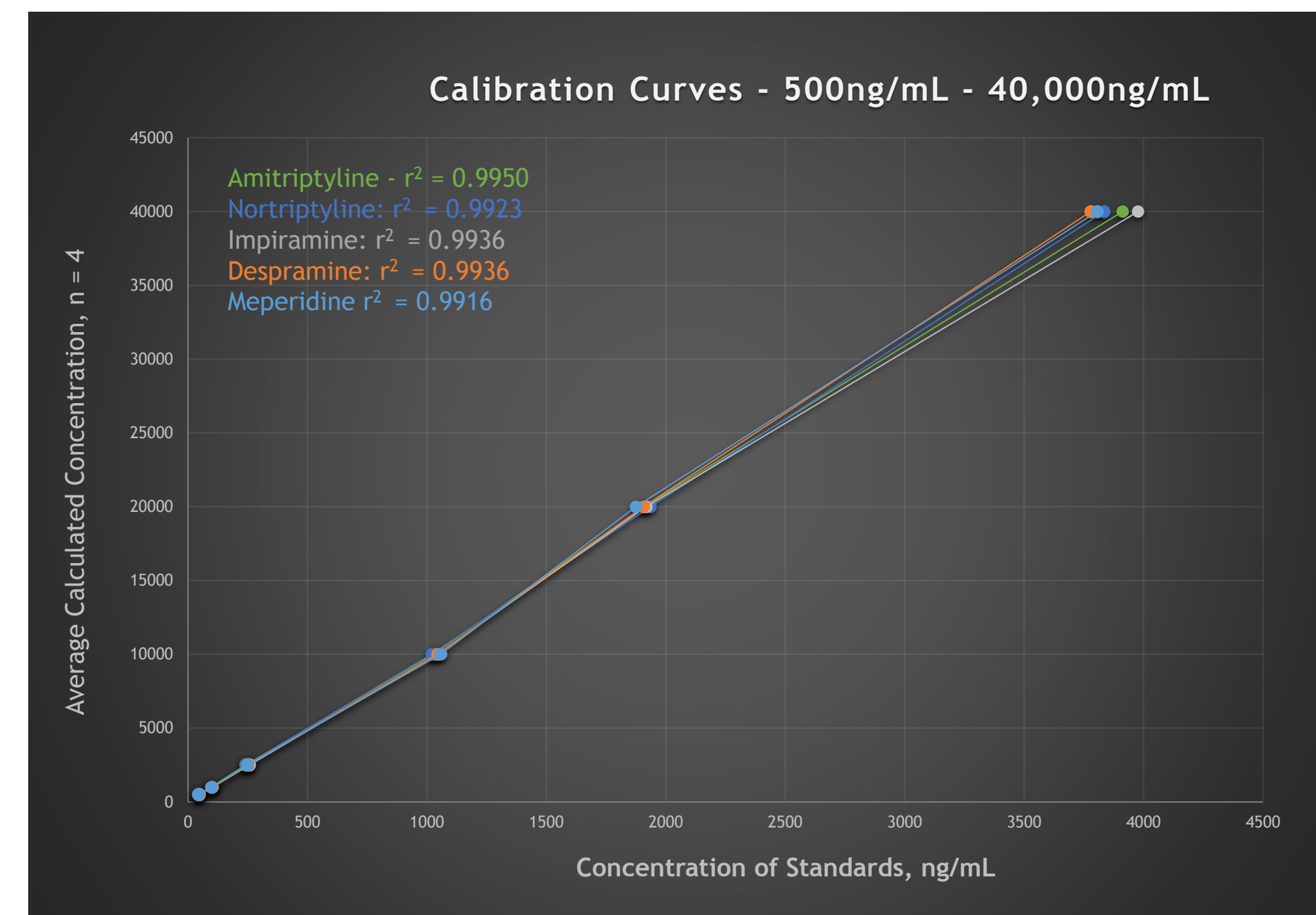


Fig. 2 Calibration curves for Amitriptyline, Nortriptyline, Imipramine, Desipramine, and Meperidine. Correlation Coefficients are > 0.99.



| Amitriptyline - Linearity/Carryover | | | | | | Nortriptyline Linearity/Carryover | | | | | | Imipramine Linearity/Carryover | | | | | |
|-------------------------------------|----------------|------------------|-------------------|-------------|----------------|-----------------------------------|-------------------|-------------|----------------|------------------|-------------------|---------------------------------|------|--------|-------|------|------------|
| Sample | Conc | Mean | SD | % CV | % Accuracy | Sample | Conc | Mean | SD | % CV | % Accuracy | Sample | Conc | Mean | SD | % CV | % Accuracy |
| Level 1 | 50 | 47.5 | 2.3 | 4.8 | 95.0 | Level 1 | 50 | 45.4 | 9.9 | 21.9 | 90.9 | Level 1 | 50 | 47.3 | 5.8 | 12.2 | 94.6 |
| Level 2 | 100 | 96.2 | 14.5 | 15.0 | 96.2 | Level 2 | 100 | 99.8 | 10.1 | 10.2 | 99.8 | Level 2 | 100 | 102.6 | 13.8 | 13.5 | 102.6 |
| Level 3 | 250 | 241.1 | 21.0 | 8.7 | 96.5 | Level 3 | 250 | 244.9 | 29.7 | 12.1 | 98.0 | Level 3 | 250 | 257.7 | 26.2 | 10.2 | 103.1 |
| Level 4 | 1000 | 1020.1 | 70.0 | 6.8 | 102.9 | Level 4 | 1000 | 1018.9 | 75.2 | 7.4 | 101.9 | Level 4 | 1000 | 1046.2 | 46.5 | 4.4 | 104.4 |
| Level 5 | 2000 | 1908.2 | 138.7 | 7.3 | 95.4 | Level 5 | 2000 | 1935.8 | 94.1 | 4.9 | 96.8 | Level 5 | 2000 | 1918.2 | 141.2 | 7.4 | 95.9 |
| Level 6 | 4000 | 3913.0 | 193.7 | 5.0 | 97.8 | Level 6 | 4000 | 3835.2 | 210.7 | 5.5 | 95.9 | Level 6 | 4000 | 3977.4 | 251.5 | 6.3 | 99.4 |
| Blank | 0 | 0 | 0 | 0 | 0 | Blank | 0 | 0 | 0 | 0 | 0 | Blank | 0 | 0 | 0 | 0 | 0 |
| Correlation Coefficient: 0.9950 | | | | | | Correlation Coefficient: 0.9923 | | | | | | Correlation Coefficient: 0.9936 | | | | | |
| Within Run Precision | | | | | | Within Run Precision | | | | | | Within Run Precision | | | | | |
| Sample | Conc | Mean | SD | % CV | % Accuracy | Sample | Conc | Mean | SD | % CV | % Accuracy | Sample | Conc | Mean | SD | % CV | % Accuracy |
| LOD/LOQ | 50 | 47.5 | 2.3 | 4.8 | 95.0 | LOD/LOQ | 50 | 45.4 | 9.9 | 21.9 | 90.9 | LOD/LOQ | 50 | 47.3 | 5.8 | 12.2 | 94.6 |
| Recovery | | | | | | Recovery | | | | | | Recovery | | | | | |
| Sample | Mean Extracted | Mean Unextracted | % Recovery | Sample | Mean Extracted | Mean Unextracted | % Recovery | Sample | Mean Extracted | Mean Unextracted | % Recovery | | | | | | |
| L1 | 6893 | 38402 | 17.7 | L1 | 18724 | 32597 | 57.4 | L1 | 65777 | 32367.75 | 20.4 | | | | | | |
| Ion Suppression | | | | | | Ion Suppression | | | | | | Ion Suppression | | | | | |
| Sample | Mean Extracted | Mean Unextracted | % Ion Suppression | Sample | Mean Extracted | Mean Unextracted | % Ion Suppression | Sample | Mean Extracted | Mean Unextracted | % Ion Suppression | | | | | | |
| L1 Standard | 163276.7 | 398669.7 | 59 | L1 Standard | 240252.7 | 395191.0 | 27 | L1 Standard | 146690.0 | 328303.3 | 55 | | | | | | |
| ISTD | 1339615.0 | 394045.0 | 66 | ISTD | 1339615.0 | 394045.0 | 66 | ISTD | 232738.3 | 627660.3 | 37 | | | | | | |
| Desipramine Linearity/Carryover | | | | | | Meperidine Linearity/Carryover | | | | | | | | | | | |
| Sample | Conc | Mean | SD | % CV | % Accuracy | Sample | Conc | Mean | SD | % CV | % Accuracy | | | | | | |
| Level 1 | 50 | 45.2 | 7.1 | 15.7 | 90.3 | Level 1 | 50 | 43.0 | 10.2 | 23.8 | 86.0 | | | | | | |
| Level 2 | 100 | 101.7 | 11.3 | 11.1 | 101.7 | Level 2 | 100 | 99.4 | 10.4 | 10.5 | 99.4 | | | | | | |
| Level 3 | 250 | 250.9 | 21.8 | 8.7 | 100.4 | Level 3 | 250 | 249.6 | 27.3 | 11.0 | 99.8 | | | | | | |
| Level 4 | 1000 | 1044.5 | 83.0 | 7.9 | 104.5 | Level 4 | 1000 | 1058.3 | 81.8 | 7.7 | 105.8 | | | | | | |
| Level 5 | 2000 | 1907.2 | 131.4 | 6.9 | 95.4 | Level 5 | 2000 | 1874.7 | 19.8 | 1.1 | 93.7 | | | | | | |
| Level 6 | 4000 | 3779.8 | 288.5 | 7.6 | 94.5 | Level 6 | 4000 | 3806.5 | 176.1 | 4.6 | 95.2 | | | | | | |
| Blank | 0 | 0 | 0 | 0 | 0 | Blank | 0 | 0 | 0 | 0 | 0 | | | | | | |
| Correlation Coefficient: 0.9926 | | | | | | Correlation Coefficient: 0.9916 | | | | | | | | | | | |
| Within Run Precision | | | | | | Within Run Precision | | | | | | | | | | | |
| Sample | Conc | Mean | SD | % CV | % Accuracy | Sample | Conc | Mean | SD | % CV | % Accuracy | | | | | | |
| LOD/LOQ | 50 | 45.2 | 7.1 | 15.7 | 90.3 | LOD/LOQ | 50 | 43.0 | 10.2 | 23.8 | 86.0 | | | | | | |
| Recovery | | | | | | Recovery | | | | | | | | | | | |
| Sample | Mean Extracted | Mean Unextracted | % Recovery | Sample | Mean Extracted | Mean Unextracted | % Recovery | | | | | | | | | | |
| L1 | 115437 | 187828.5 | 61.5 | L1 | 33794 | 55726 | 60.6 | | | | | | | | | | |
| Ion Suppression | | | | | | Ion Suppression | | | | | | | | | | | |
| Sample | Mean Extracted | Mean Unextracted | % Ion Suppression | Sample | Mean Extracted | Mean Unextracted | % Ion Suppression | | | | | | | | | | |
| L1 Standard | 138464.7 | 191936.7 | 28 | L1 Standard | 40364.7 | 58803.0 | 28 | | | | | | | | | | |
| ISTD | 4593416.7 | 7439159.0 | 38 | ISTD | 3049026.7 | 4927172.0 | 38 | | | | | | | | | | |

Fig. 3 Calibration curves for Ritalinic Acid, Tapentadol, Tapentadol-O-Sulfate, and Tramadol. Correlation Coefficients are > 0.99.

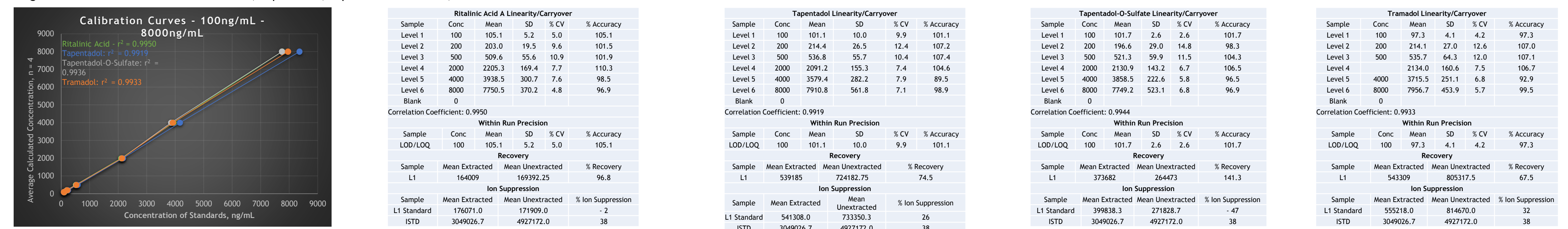


Fig. 4 Calibration curves for Cycloenzaprine, Duloxetine, Normeperidine. Correlation Coefficients are > 0.99.

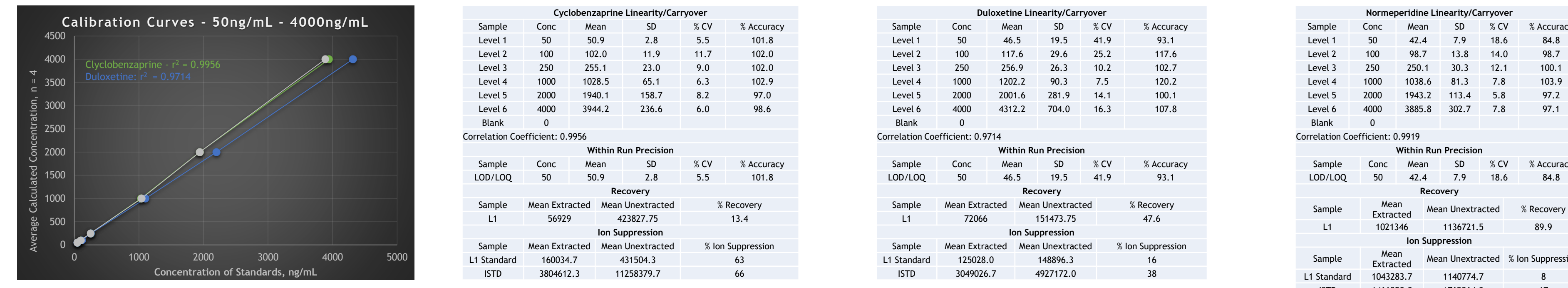


Fig. 5 Calibration curves for Gabapentin and Pregabalin. Correlation Coefficients are > 0.99.

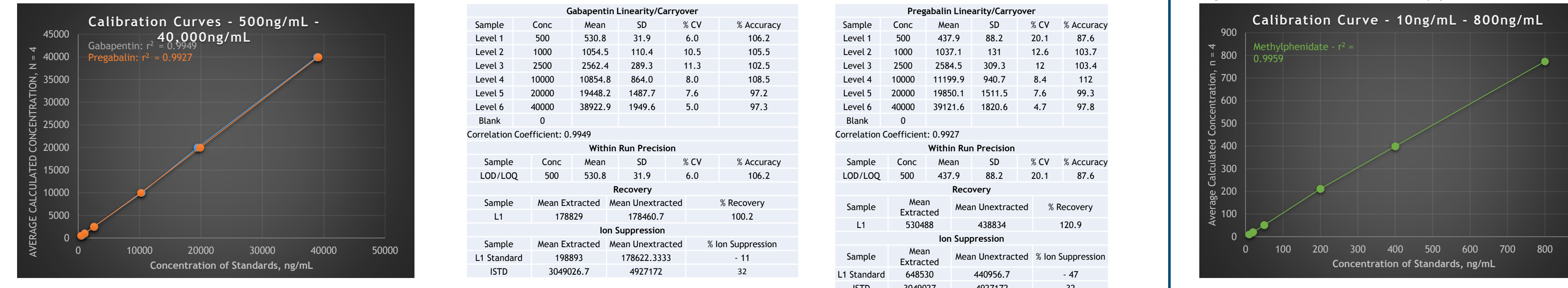
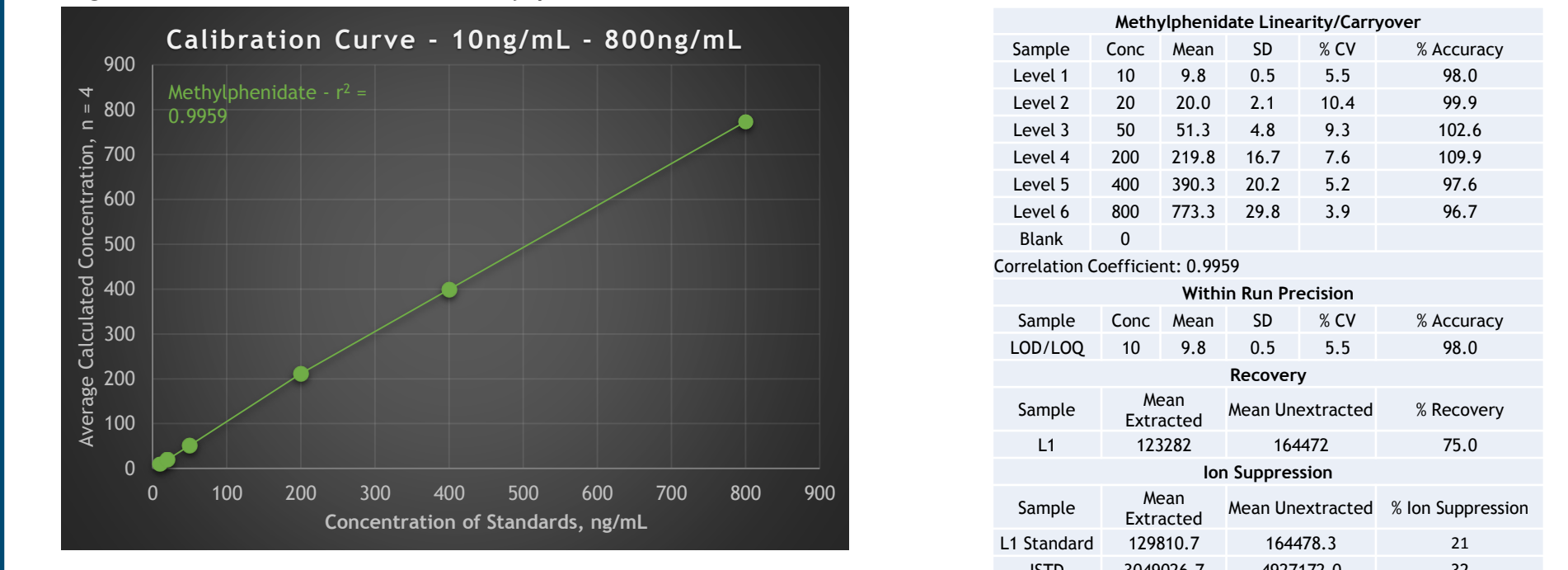


Fig. 6 Calibration curve for Methylphenidate. Correlation Coefficient is > 0.99.



Conclusion

This validated method alleviates the need for sample clean-up by SPE or SLE thereby reducing the amount of equipment required, solvent usage and sample preparation time. Samples are filtered by pipetting the sample into the filter vial shell, inserting the plunger into the shell, and then pushing the plunger into the shell. The filtration process from sample pipetting to autosampler ready only requires 15 seconds. Benefits to the use of Thomson eXtreme|FV® include lower cost, faster sample preparation time, less use and disposal of organic solvents.

